Functional Polymorphisms of the Cyclooxygenase-2 Gene and Risk for Chronic Obstructive Pulmonary Disease in Japanese Population

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Chronic obstructive pulmonary disease (COPD) is characterized by chronic inflammation. Cyclooxygenase-2 (COX-2) plays a very important role in the progress of inflammation. Several potentially functional polymorphisms of the COX-2 gene were identified and proposed to be associated with COPD susceptibility. In this study, we aimed to investigate a possible association between COX-2 (-765G>C; -1195G>A) polymorphisms and the risk of COPD in the Japanese population. COX-2 gene polymorphisms were identified by polymerase chain reaction-restriction fragment-length polymorphism (PCR-RFLP) in 103 Japanese patients with COPD and 129 healthy controls. The distributions of genotypes and alleles were compared by using the χ^2 test. The odds ratios (OR) and 95% confidence intervals (95%CI) were calculated by a logistic regression analysis. The frequencies of the GG, GC, CC genotypes of COX-2-765G>C and GG, GA, AA genotypes of COX-2-1195G>A in the COPD group were 94.2%, 5.8%, 0.0% and 14.6%, 40.8%, 44.6%, respectively. These frequencies in control group were 89.1%, 10.9%, 0.0% and 23.3%, 44.9%, 31.8%, respectively. The COX-2-1195AA was more prevalent in the COPD group (44.6%) than the control group (31.8%). The distribution of the -765C was higher in the control group (5.5%) in comparison to the

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COPD group (2.9%). In conclusion, the COX-2-1195AA genotype was associated with an increased risk for COPD in the Japanese population.

Key words: Japanese, COX-2, polymorphisms, risk of COPD

INTRODUCTION

Chronic obstructive pulmonary disease (COPD) is a major cause of morbidity and mortality worldwide [1]. The World Health Organization estimates that more than 3 million people died of COPD in 2005, which corresponds to 5% of all deaths globally [2]. COPD is an inflammatory disease characterized by the progressive deterioration of pulmonary function and increasing airway obstruction. Accumulating studies show several genetic risk factors are associated with susceptibility to COPD, such as α 1-antitrypsin, α 1-antichymotrypsin, cystic fibrosis transmembrane regulator, vitamin D-binding protein, α 2-macroglobulin, cytochrome P4501A1, blood group antigens, human leukocyte antigen, immunoglobulin deficiency, and haptoglobin [3].

Cyclooxygenase (COX) is a key enzyme in the conversion of arachidonic acid into prostaglandins. COX-2 is an induced isoform which is characteristic of cells involved in inflammatory processes [4]. The over-expression of COX-2 is linked to enhancement of cellular proliferation, promotion of angiogenesis, inhibition of apoptosis, stimulation of invasion, and suppression of immune responses [5, 6]. Several common single nucleotide polymorphisms (SNPs) of the COX-2 gene are reportedly functional and associated with human tumors of various organs [7-13]. On the other hand, a few studies have so far suggested that COX-2 plays a biologically important role in the inflammation of respiratory disease [14-16].

The purpose of this study was to evaluate whether the COX-2 -765G>C or -1195G>A polymorphisms are associated with susceptibility to COPD.

MATERIALS AND METHODS

Patients and Samples

One hundred and three patients with COPD and 129 healthy controls took part in this study. Patients were recruited from the Department of Internal Medicine, Division of Clinical Oncology & Respiratory Medicine, Shimane University Hospital (Shimane, Japan). COPD was diagnosed based on clinical history, physical examination, and pulmonary functional tests, according to the GOLD criteria [1]. The patients had an FEV1/FVC ratio of <70% and FEV1 of <80% of predicted values. The control subjects were sex- and age-matched, and visited the same hospitals for a routine health checkup. Control subjects had normal pulmonary function tests (FEV1/ FVC > 70% and FEV1 > 80% of predicted values) with no history of airway disease or abnormal findings on chest radiography.

All subjects gave written informed consent for participation in the study. The study protocol was approved by local ethics committee.

DNA preparation and genotype determination

Peripheral blood samples were collected from each study subject into EDTA-containing tubes. Genomic DNA was extracted from peripheral whole blood samples using a DNA extractor WB-Rapid kit (Wako, Osaka, Japan). PCR-RFLP was used for the analysis of COX-2 polymorphisms. The PCR was carried out in a reaction mixture of 50 uL containing 2.5 U Taq and 1 uL template DNA with a concentration of approximately 50-150 ng/ml. The COX-2-765G>C polymorphism was determined using primers 5'-ATT CTG GCC ATC GCC GCT TC-3' (forward) and 5'-CTC CTT GTT TCT TGG AAA GAG ACG -3' (reverse). The -1195G>A polymorphism was determined using primers 5'- CCC TGA GCA CTA CCC ATG AT-3' (forward) and 5'-GCC CTT CAT AGG AGA TAC TGG-3' (reverse). The PCR cycling conditions for the -765G>C and -1195G>A polymorphisms analyses were: 5 min at 96°C followed by 32 cycles of 20 sec at 96°C , 30 sec at 52°C , and 30 sec at 72°C , with a final extension of 72°C for 6 min. The 157 bp PCR product of -765G>C was digested using Bsh1236I into two fragments of 134 bp and 23 bp for the 765G allele. The 273 bp product of the -1195G>A PCR product was digested with PvuII into 220 bp and 53 bp fragments for the -1195G allele. The digested products were separated by electrophoresis in 2% agarose gels stained with ethidium bromide under UV light (Figs 1 and 2). The PCR products were sequenced directly at random to rule out the possibility of PCR errors.



Fig. 1. Patterns of PCR-RFLP for COX-2-765G>C. M: 100 bp DNA ladder; Line 1, 2, 3, 5: Homozygous -765GG genotype; Line 4: Heterozygous -765GC genotype; Line 6: Positive control of homozygous-765CC genotype; Line 7: Positive control of homozygous -765GG genotype. The PCR product was digested with *Bsh12361*, which cleaved wild genotype G into 134 bp and 23 bp fragments, variant allele C remained an uncleaved 157 bp fragment.



Fig. 2. Patterns of PCR-RFLP for COX-2-1195G>A. M: 100 bp DNA ladder; Line 1, 2, 3: Homozygous-1195GG genotype; Line 4, 5, 6,7: Homozygous-1195AA genotype; Line 8, 9, 10,11: Heterozygous-1195GA genotype. The PCR product was digested with *PvuII*, which cleaved wild genotype G into 220 bp and 53 bp fragments, variant allele A remained an uncleaved 273 bp fragment.

Statistical analysis

The Statistical Package for Social Sciences software package (Version 17.0) was used for all statistical analyses. Data are expressed as the mean \pm standard deviation (SD) or numbers (%) of subjects. The allele and genotype frequencies of the patients and controls were compared with the χ^2 test. The analysis of association between COX-2 polymorphisms and COPD was based on the 95% CI for the disease odds ratio (OR) calculated. Logistic regression was used to assess the association between the genotype and the risk of COPD. The Hardy-Weinberg equilibrium was assessed for the distributions of genotype in each group. *P* <0.05 was considered to be statistically significant.

RESULTS

Baseline clinical characteristics of study subjects

Table 1 summarizes the demographic characteristics and relevant clinical parameters of all study subjects. The age and gender distributions were similar between COPD and control groups. Nonsmokers were more in controls than COPD group. As expected, the FEV1, % pred and FEV1/FVC, % in the controls were higher than those in the COPD group.

Analysis of COX-2-765G > C and -1195G > A genotypes

Table 2 summarizes the SNP distributions and allele frequencies. All genotypic distributions accorded with the Hardy-Weinberg equilibrium (P > 0.05).

The frequencies of the COX-2-765GG, GC, and CC genotypes among the COPD group were 94.2%, 5.8%, and 0.0%, respectively, in comparison to 89.1%, 10.9%, and 0.0%, respectively, in the controls. The -765C allele was quite scarce, with no CC homozygote in either COPD or control subjects. The GC genotype was lower in COPD than control subjects (5.8% versus 10.9%), but the decreased risk was not significant (adjusted OR=0.509, 95%CI 0.181-1.376, P = 0.223).

The frequencies of the COX-2-1195GG, GA, and AA genotypes in COPD were 14.6%, 40.8%, and 44.6%, respectively, in comparison to 23.3%, 44.9%, and 31.8%, respectively, in the controls. The COX-2-1195G>A polymorphism was more general, with similar distributions of -1195GG and GA genotypes between the two groups, but a higher frequency of the -1195AA homozygote in COPD subjects than controls (44.6% versus 31.8%). The individuals who had the -1195AA genotype had an increased risk for COPD (adjusted OR=2.246, 95%CI 1.064-4.759, P=0.041).

Table 1. Baseline clinical characteristics of study subjects.

	COPD (n=103)	Controls (n=129)	P value
Age	63.5 ± 6.2	58.4 ± 9.1	0.420
Gender			
М	100 (97.1)	125 (92.4)	
F	3 (2.9)	4 (7.6)	1.000
Smoking			
No	2 (1.9)	33 (25.6)	
Yes	101 (98.1)	96 (74.4)	< 0.001
FEV1, %pred	51.4 ± 0.9	99.0 ± 1.4	< 0.001
FEV1/FVC, %	48.8 ± 1.1	85.4 ± 1.7	< 0.001

Values represent the number (%) of subjects or the mean \pm SD.

FEV1, forced expiratory volume in 1 second; FVC, forced vital capacity.

SNP	COPD (n=103)	control (n=129)	P value	OR (95%CI)	OR* (95%CI)	P value
765G>C						
GG	97 (94.2)	115 (89.1)		Reference	Reference	
GC	6 (5.8)	14 (10.9)	0.240	0.508 (0.188-1.373)	0.509 (0.181-1.376)	0.223
CC	0 (0.0)	0 (0.0)	NS			
Allele						
G	97.1%	94.5%		Reference	Reference	
С	2.9%	5.5%	0.498	0.490 (0.119-2.015)	0.478 (0.126-1.988)	0.445
HWE $(x2/P)$	0.10/0.97	0.57/0.95				
1195G>A						
GG	15(14.6)	30(23.3)		Reference	Reference	
GA	42 (40.8)	58(44.9)	0.362	1.448 (0.694-3.024)	1.441 (0.690-3.029)	0.357
AA	46 (44.6)	41 (31.8)	0.043	2.244 (1.061-4.747)	2.246 (1.064-4.759)	0.041
Allele						
G	35.0%	45.8%		Reference	Reference	
А	65.0%	54.2%	0.150	1.582 (0.896-2.794)	1.577 (0.885-2.808)	0.147
HWE (<i>x</i> 2/ <i>P</i>)	0.86/0.65	0.63/0.66				

Table 2. COX-2 genotype distributions in patients with COPD and controls.

HWE, Hardy-Weinberg equilibrium test; OR, odds ratio; 95%CI, 95% confidence interval;

*Adjusted for age, gender and smoking history.

NS, not significant.

DISCUSSION

The present study is the first one to assess the role of the COX-2 gene polymorphisms in COPD in the Japanese population. There was no significant difference in the genotype distribution and the allele frequency of COX-2-765G>C among the COPD and control groups. The -765GG genotype was more prevalent in the Japanese population, the frequency of -765GG genotype was 94.2% in COPD patients and 89.1% in controls. No -765CC homozygosity was found in both groups. The distribution of the -765C allele was more in controls than COPD subjects (5.5% versus 2.9%), but the increase was not statistically significant (adjusted OR=0.478, 95%CI 0.126-1.988, P=0.445). The reported frequency of -765C allele seems to vary among different ethnic populations, it is 21%-32% in Caucasian and African Americans [17], 14%-28% in Europeans [18-21], 16% in healthy Hindus [22], 5% in Singaporeans [9],

2.3% in Japanese [23]. In our study, the frequency of C allele was greater than the reported in Japanese, as described above, the discrepancy could be explained by diverse geographical area. Indeed, the complex mechanisms of genovariation are unclear, and the effects of external factors could not be excluded. There are many studies about the effect of COX-2-765CC genotype or -765C allele, but the conclusions are conflicting. For instance, individuals carrying the -765C allele have lower risks of myocardial infarction and ischemic stroke in Italy [17], and have a decreased risk of Alzheimer's disease in the USA [24], while another study showed the -765CC genotype increases the risk of asthma and the PGE2 production is 10 times higher than the GG group [20]. These outcomes indicate that the COX-2-765G>C polymorphism may have a significantly different modulating effects on various disease phenotypes in different races and regions.

The current study was also the first that addressed the correlation between COPD and COX-2-1195G>A in the Japanese population. Individuals carrying -1195AA genotype had an increased risk for COPD. Polymorphism in COX-2-1195G>A are fairly common in humans. The distributions of -1195GG, GA and allele G, A were similar in the COPD and control groups, but the -1195AA genotype was higher in COPD subjects than controls (44.6% versus 31.8%), suggesting that the -1195AA was a risk genotype for COPD (P=0.041, adjusted OR=2.246, 95%CI 1.064-4.759). A possible mechanism for this effect is that the -1195A allele creates a c-Mvb binding site to upregulate the gene expression, thus the -1195A allele is a risk factor by increasing the promoter activity; this possibility is consistent with other studies [8, 10, 25-27].

In conclusion, the current findings suggested that COPD was not associated with the COX-2-765G>C polymorphism, but not the COX-2-1195G>A polymorphism, the -1195AA genotype was a risk factor for COPD. Further studies with a larger sample size are necessary to clarify the role of COX-2 in COPD among different geographic areas and races.

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REFERENCES

- 1) GOLD -Global Initiative For Chronic Obstructive Lung Disease – UPDATE (2007). "Global Strategy for the Diagnosis, Management, and prevention of Chronic Obstrutive Pulmonary Disease." NHLBI/WHO. http://www.goldcopd.com. Acessed in June 2011.
- 2) World Health Organization. Chronic respiratory diseases. Available at: http://www.who.int/respiratory/copd/burden/en/index.html.

- 3) Sandford AJ, Weir TD and Pare PD (1997) Genetic risk factors for chronic obstructive pulmonary disease. *Eur Respir J* 10: 1380-1391.
- 4) Minghetti L (2004) Cyclooxygenase-2 (COX-2) in inflammatory and degenerative brain disease. *J Neuropathol Exp Neurol* 63: 901-910.
- 5) Wang D and Dubois RN (2006) Prostaglandins and cancer. *Gut* 55: 115-122.
- 6) Gasparini G, Longo R and Morabito A (2003) Inhibitors of cyclo-oxygenase 2: A new class of anticancer agents? *Lancet Oncol* 4: 605-615.
- 7) Ferguson HR, Wild CP, Anderson LA, Murphy SJ, Johnston BT, Murray LJ, Watson RG, McGuigan J, Reynolds JV and Hardie LJ (2008) Cyclooxygenase-2 and inducible nitric oxide synthase gene polymorphisms and risk of reflux esophagitis, Barrett's esophagus, and esophageal adenocarcinoma. *Cancer Epidemiol Biomark Prevent* 17: 727-731.
- 8) Zhang X, Miao X, Tan W, Ning B, Liu Z, Hong Y, Song W, Guo Y, Zhang X, Shen Y, Qiang B, Kadlubar FF and Lin D (2005) Identification of functional genetic variants in cyclooxygenase-2 and their association with risk of esophageal cancer. *Gastroenterology* 129: 565-576.
- 9) Koh WP, Yuan JM, van den Berg D, Lee HP and Yu MC (2004) Interaction between cyclooxygenase-2 gene polymorphism and dietary n-6 polyunsaturated fatty acids on colon cancer risk: the Singapore Chinese Healthy Study. *Br J Cancer* 90: 1760-1764.
- 10) Liu F, Pan K, Zhang X, Zhang Y, Zhang L, Ma J, Dong C, Shen L, Li J, Deng D, Lin D and You W (2006) Genetic variants in cyclooxyygenase-2: expression and risk of gastric cancer and its precursors in a Chinese population. *Gastroenterol* 130: 1975-1984.
- Hosomi Y, Yokose T, Hirose Y, Nakajima R, Nagai K, Nishiwaki Y and Ochiai A (2000) Increased cyclooxygenase 2 expression occurs frequently in precursor lesions of human adenocarcinoma of the lung. *Lung cancer* 30: 73-81.
- 12) Yoshimatsu K, Altorki NK, Golijanin D, Zhang F, Jakobsson PJ, Dannenberg AJ and Subbaramaiah K (2001) Inducible prostaglandin E synthase is overexpressed in non-small cell lung

cancer. Clin Cancer Res 7: 2669-2674.

- 13) Gao J, Ke Q, Ma HX, Wang Y, Zhou Y, Hu ZB, Zhai XJ, Wang XC, Qing JW, Chen WS, Jin GF, Liu JY, Tan YF, Wang XR and Shen HB (2007) Functional polymorphisms in the cy-clooxygenase 2 (COX-2) gene and risk of breast cancer in a Chinese population. *J Toxicol Environ Health A* 70: 908-915.
- 14) Shi J, Misso NL, Kedda MA, Horn J, Welch MD, Duffy DL, Williams C and Thompson PJ (2008) Cyclooxygenase-2 gene polymorphisms in an Australian population: association of the -1195G>A promoter polymorphism with mild asthma. *Clin and Exp Allergy* 38: 913-920.
- 15) Lin CC, Lee IT, Yang YL, Lee CW, Kou YR and Yang CM (2010) Induction of COX-2/ PGE2/IL-6 is crucial for cigarette smoke extractinduced airway inflammation: Role of TLR4-dependent NADPH oxidase activation. *Free Radical Bio Med* 48: 240-254.
- 16) Li H, Bradbury JA, Dackor RT, Edin ML, Graves JP, DeGraff LM, Wang PM, Bortner CD, Maruoka S, Lih FB, Cook DN, Tomer KB, Jetten AM and Zeldin DC (2011) Cyclooxygenase-2 regulates Th17 cell differentiation during allergic lung inflammation. *Am J Respir Crit Care Med* 1: 37-49.
- 17) Pereira C, Sousa H, Ferreira P, Fragoso M, Moreira-Dias L, Lopes C, Medeiros R and Dinis-Ribeiro M (2006) -765G>C COX-2 polymorphism may be a susceptibility marker for gastric adenocarcinoma in patients with atrophy or intestinal metaplasia. *World J Gastroenterol* 12: 5473-5478.
- 18) Papafili A, Hill MR, Brull DJ, McAnulty RJ, Marshall RP, Humphries SE and Laurent GJ (2002) Common promoter variant in cyclooxyygenase-2 represses gene expression: evidence of role in acute-phase inflammation response. *Arterioscler Thromb Vasc Biol* 22: 1631-1636.
- 19) Cipollone F, Toniato E, Martinotti S, Fazia M, Iezzi A, Cuccurullo C, Pini B, Ursi S, Vitullo G, Averna M, Arca M, Montali A, Campagna F, Ucchino S, Spigonardo F, Taddei S, Virdis

A, Ciabattoni G, Notarbartolo A, Cuccurullo F and Mezzetti A (2004) A polymorphism in the cyclooxygenase 2 gene as an inherited protective factor against myocardial infarction and stroke. *JAMA* 291: 2221-2228.

- 20) Szczklik W, Sanak M and Seceeklik A (2004) Functional effects and gender association of COX-2 gene polymorphism G-765C in bronchial asthma. *J Allergy Clin Immunol* 114: 248-253.
- 21) Shi J, Misso NL, Duffy DL, Thompson PJ and Kedda MA (2004) A functional polymorphism in the promoter region of the cyclooxygenase-2 gene is not associated with asthma and atopy in an Australian population. *Clin Exp Allergy* 34: 1714-1718.
- 22) Arif E, Vibhuti A, Deepak D, Singh B, Siddiqui MS and Pasha MA (2008) COX2 and P53 risk-alleles coexist in COPD. *Clin Chim Acta* 397: 48-50.
- 23) Hamajima N, Takezaki T, Matsuo K, Saito T, Inoue M, Hirai T, Kato T, Ozeki J and Tajima K (2001) Genotype frequencies of cyclooxygenase 2 (COX-2) rare polymorphisms for Japanese with and without colorectal cancer. *Asian Pac J Cancer Prev* 2: 57-62.
- 24) Abdullah L, Ait-Ghezala G, Crawford F, Crowell TA, Barker WW, Duara R and Mullan M (2006) The cyclooxygenase 2-765C promoter allele is a protective factor for Alzheimer's disease. *Neurosci Lett* 395: 240-243.
- 25) Xie CJ, Xiao LM, Fan WH, Xuan DY and Zhang JC (2009) Common single nucleotide polymorphisms in cyclooxygenase-2 and risk of severe chronic periodontitis in a Chinese population. *J clin Periodontol* 36: 198-203.
- 26) Ramsay RG, Friend A, Vizantios Y, Freeman R, Sicurella C, Hammett F, Armes J and Venter D (2000) Cyclooxyygenase-2,a colorectal cancer nonsteroidal anti-inflammatory drug target, is regulated by cMYB. *Cancer Res* 60: 1805-1809.
- 27) Ramsay RG, Barton AL and Gonda TJ (2003) Targeting c-MYB expression in human disease. *Expert Opin Ther Target* 7: 235-248.