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Abstract: [Introduction]

An increasing number of adult patients have been diagnosed with fatty acid β -oxidation disorders with the increase of diagnostic technologies. In this study, clinical, biochemical, and molecular characteristics in 2 Japanese patients with the adult-onset form of glutaric acidemia type II (GA2) were investigated, and compared with those of pediatric cases.

[Methods]

The patients were a 58-year-old male and a 31-year-old male. In both cases, episodes of myopathic symptoms, including myalgia, muscle weakness, and liver dysfunction of unknown causes, had been noted for the past several years. Muscle biopsy, urinary organic acid analysis (OA), blood acylcarnitine (AC) analysis, immunoblotting, genetic analysis, and an in vitro probe acylcarnitine (IVP) assay were used for diagnosis and investigation.

[Results]

In both cases, there was no obvious abnormality in blood ACs or urinary OA, while there was a slight increase in medium- and long-chain ACs in serum; also, fat deposits were observed in the muscle biopsy. Immunoblotting and gene analysis revealed that both patients had GA2 due to a defect of ETFDH. The IVP assay presented with no special abnormalities in both cases.

[Conclusion]

The late-onset GA2 is separated between the infantile-onset and myopathic forms. In the myopathic form, episodic muscular symptoms or liver dysfunction are mainly exhibited after later childhood. Muscle biopsy and serum (or plasma) AC analysis is a clue to reach the correct diagnosis, because other biochemical tests, such as blood AC analysis, urinary OA, or IVP assay, show less abnormality in the myopathic form compared with the infantile-onset form.

Suggested Reviewers: Toshiyuki Fukao MD, PhD

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Professor Kure is well known as expert in genetics and inherited metabolic disease.

Opposed Reviewers:

Dr. Masashi Mizuguchi
Editor-in-Chief
Brain and Development

July 17, 2015

Dear Dr. Mizuguchi:

I am pleased to submit an article entitled “Clinical, biochemical and molecular investigation of adult-onset glutaric acidemia type II: characteristics in comparison with pediatric cases” by Kenji Yamada, Hironori Kobayashi, Ryosuke Bo, Tomoo Takahashi, Jamiyan Purevusren, Yuki Hasegawa, Takeshi Taketani, Seiji Fukuda, Takuya Ohkubo, Yokota Takanori, Mutsufusa Watanabe, Taiji Tsunemi, Hidehiro Mizusawa, Hiroshi Takuma, Ayako Shioya, Akiko Ishii, Akira Tamaoka, Yosuke Shigematsu, Hideo Sugie, and Seiji Yamaguchi for consideration for publication as an original basic research article in *Brain and Development*.

In this study, we assessed the clinical, biochemical and molecular characteristics of 2 Japanese patients with adult-onset glutaric acidemia type II (GA2) using techniques such as muscle biopsy, urinary organic acid analysis, serum acylcarnitine (AC) assays, immunoblotting and genetic analysis. Both patients had GA2 due to defective electron-transferring-flavoprotein dehydrogenase (ETFDH); additionally, medium- and long-chain ACs were increased in their sera, and fat deposits were observed in their muscle. Thus, muscle biopsy and serum AC analysis are useful to correctly diagnose adult-onset GA2. In addition, fewer biochemical abnormalities are seen in adult-onset GA2 compared with pediatric cases.

We believe that this manuscript is appropriate for publication in *Brain and Development* because it provides novel diagnostic criteria for identifying the adult-onset form of GA2, which is useful for newborn screening. Our findings will be of interest to clinicians and researchers alike aiming to accurately diagnose GA2 in adults to improve patient outcomes.

This manuscript has not been published and is not under consideration for publication elsewhere. We have no conflicts of interest to disclose. All authors have read and approved the final version of the manuscript. Thank you for your consideration, and we look forward to hearing from you at your earliest convenience.

Sincerely,

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1 **Clinical, biochemical and molecular investigation of adult-onset glutaric acidemia**
2 **type II: characteristics in comparison with pediatric cases**

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1
2 **1 Abstract**
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5 2 [Introduction] An increasing number of adult patients have been diagnosed with fatty
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7 3 acid β -oxidation disorders with the rising use of diagnostic technologies. In this study,
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9 4 clinical, biochemical, and molecular characteristics of 2 Japanese patients with
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11 5 adult-onset glutaric acidemia type II (GA2) were investigated and compared with those
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13 6 of pediatric cases.
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19 8 [Methods] The patients were a 58-year-old male and a 31-year-old male. In both cases,
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21 9 episodes of myopathic symptoms, including myalgia, muscle weakness, and liver
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23 10 dysfunction of unknown cause, had been noted for the past several years. Muscle biopsy,
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25 11 urinary organic acid analysis (OA), blood acylcarnitine (AC) analysis, immunoblotting,
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27 12 genetic analysis, and an *in vitro* probe acylcarnitine (IVP) assay were used for diagnosis
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29 13 and investigation.
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36 15 [Results] In both cases, there was no obvious abnormality in blood AC or urinary OA,
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38 16 although there was a slight increase in medium- and long-chain ACs in serum; also, fat
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40 17 deposits were observed in the muscle biopsy. Immunoblotting and gene analysis
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42 18 revealed that both patients had GA2 due to a defect in electron transfer flavoprotein
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44 19 dehydrogenase (ETFDH). The IVP assay indicated no special abnormalities in either
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46 20 case.
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53 22 [Conclusion] Late-onset GA2 is separate from the infantile-onset and myopathic forms.
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55 23 In the myopathic form, episodic muscular symptoms or liver dysfunction are primarily
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57 24 exhibited after later childhood. Muscle biopsy and serum (or plasma) AC analysis allow
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1 accurate diagnosis in contrast with other biochemical tests, such as blood AC analysis,
2 urinary OA, or the IVP assay, which show fewer abnormalities in the myopathic form
3 versus the infantile-onset form.

4
5 **Keywords**

6 multiple acyl-CoA dehydrogenase deficiency (glutaric acidemia type II), adult onset,
7 myopathy, serum acylcarnitine, immunoblotting, *in vitro* probe acylcarnitine assay

8

1 Introduction

Many organic acidemias or fatty acid oxidation disorders (FAODs) are often believed to develop in childhood, especially in early infancy [1]. However, an increasing number of adult patients with inherited metabolic diseases (IMDs) has recently been identified with new developments in diagnostic technologies, including mass spectrometry, and the spread of knowledge regarding IMDs, even in the field of adult neurology.

Glutaric acidemia type II (GA2) is an autosomal recessive disease caused by a defect in electron transfer flavoprotein (ETF) or ETF dehydrogenase (ETFHDH), resulting in deficiencies in multiple acyl-CoA dehydrogenases, such as short-, medium-, and long-chain acyl CoA dehydrogenases, as well as isovaleryl-CoA dehydrogenase, glutaryl-CoA dehydrogenase, and sarcosine-CoA dehydrogenase [2, 3]. GA2 has been clinically classified into 2 types: 1) the neonatal-onset type, which develops during the neonatal period or early infancy and is often severe, and 2) the late-onset type, which develops after the infantile period [4].

Patients with the neonatal-onset type of GA2 develop severe respiratory failure, cardiomyopathy, hypotonia, metabolic acidosis, and profound hypoglycemia soon after birth, and they often have a fatal outcome in early infancy. Some patients with this type have congenital anomalies, including Potter's face or polycystic kidney disease [5, 6].

In the late-onset type, intermittent episodic attacks of lethargy, hypoglycemia, and hyperammonemia, or, occasionally, acute encephalopathy or sudden death triggered by infection, diarrhea, or long fasting are seen starting in early childhood [7-9].

Recently, several adult-onset GA2 cases have been reported [10-13]. However, it is not always easy to establish the correct diagnosis. In this study, the clinical,

1
2 1 biochemical, and pathological characteristics of 2 cases of adult-onset GA2 were
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4 2 investigated and compared with those of pediatric cases.
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4 **2 Material and methods**

5 *2.1 Patients*

6 Case 1 was a 58-year-old male with chief complaints of episodic myalgia and
7 muscle weakness. The clinical course of case 1 has been reported previously [14]. His
8 younger brother died unexpectedly from an unknown cause in his 30s. The patient
9 sometimes had general fatigue, myalgia, or muscle weakness as early as in his 40s.
10 Those symptoms progressively worsened in his 50s, and he began to use a wheelchair
11 because of persistent muscle weakness and myalgia. Furthermore, he had 3 episodes of
12 unconsciousness after the age of 50. Although he was hospitalized at the third episode,
13 there were no obvious abnormalities in routine biochemical tests, including blood sugar
14 and liver function. He visited several neurology clinics and hospitals to undergo a more
15 detailed examination. However, no abnormality was found, except for the occasional
16 elevation of aspartate aminotransferase (AST), alanine aminotransferase (ALT), lactate
17 dehydrogenase (LDH), and creatine kinase (CK). The diagnosis was “myopathy of
18 unknown cause”. Then, as he repeatedly developed liver dysfunction and
19 rhabdomyolysis, he was hospitalized at age 58 for detailed examination, including
20 muscle biopsy.

21 On admission, his level of consciousness was normal, and his vital signs and
22 intelligence were normal. No hepatosplenomegaly was noted. Muscle tenderness and
23 atrophy with mild sensory dysfunction were observed in his limbs, especially in the
24 lower limbs, as neurological findings. The deep tendon reflex was normal, and he was

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2 1 able to walk with support. In manual muscle testing, his muscle strength was level 2 for
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4 2 the deltoid and iliopsoas muscles and 3+ to 4 for other upper and lower limb muscles.
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7 3 Routine blood examination indicated the elevation of liver and muscle enzymes,
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9 4 such as AST (197 IU/L, normal range 10-38), ALT (215 IU/L, normal 5-40), LDH
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11 5 (2,903 IU/L, normal 100-215), and CK (2,364 IU/L, normal 36-216), as shown in Table
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16 7 Case 2 was a 31-year-old male with episodic muscle weakness and myalgia
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18 8 similar to case 1. No abnormalities in his past and family history were noted. He was
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20 9 formerly a baseball player on a non-professional team, but he developed muscle
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22 10 weakness after retiring from the baseball team at 29 years of age. Then, his exertional
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24 11 muscle weakness worsened gradually, and he began to experience difficulty in his daily
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26 12 activities. Although he visited several neurology clinics or hospitals, only liver
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28 13 dysfunction of unknown cause was occasionally noted. He was hospitalized to undergo
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30 14 further examination at 31 years of age.
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34 15 His level of consciousness and his intellectual level were normal.
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36 16 Abnormalities in vital signs and hepatosplenomegaly were not observed. His patellar
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38 17 and Achilles tendon reflexes were slightly reduced, but no pathological reflex or muscle
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40 18 atrophy was observed. The results of manual muscle testing were also within the normal
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42 19 range.
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46 20 Blood examination indicated a slight elevation of liver and muscle enzymes
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48 21 (AST 71 IU/L, ALT 84 IU/L, LDH 684 IU/L, and CK 689 IU/L), although no
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50 22 abnormalities were observed in other tests.
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54 23 *2.2 Urinary organic acid analysis*

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56 24 The urinary organic acids (OAs) were analyzed using gas chromatography
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2 1 mass spectrometry (GC/MS; QP-2010 plus; Shimadzu, Kyoto, Japan) at Shimane
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4 2 University, Japan, after solvent extraction and oxime-trimethylsilyl derivatization of
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6 3 urine samples as previously described [1, 15].
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9 4 *2.3 Blood acylcarnitine analysis*

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11 5 Acylcarnitine (AC) from blood filter paper or serum was analyzed using
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13 6 tandem mass spectrometry (MS/MS) (API-3000; Applied Biosystems, Foster City, CA,
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15 7 USA) after butyl-derivatization of samples, as previously described [16, 17].
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18 8 *2.4 Histological studies*

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20 9 Muscle biopsies were performed using the rectus femoris muscle and biceps
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22 10 brachii in cases 1 and 2, respectively. The biopsied materials were frozen and
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24 11 cryostat-sectioned for Oil-Red O staining [18].
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27 12 *2.5 Cell culture*

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29 13 Skin fibroblasts were cultured in Eagle's minimal essential medium (MEM)
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31 14 (Nissui Pharmaceutical Co., Ltd., Tokyo, Japan) supplemented with 2 mmol/L
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33 15 glutamine, 10% fetal bovine serum, and 1% penicillin/streptomycin at 37°C in a
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35 16 humidified 5% CO₂/95% air incubator until confluence [19, 20].
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39 17 *2.6 Immunoblotting*

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41 18 Twenty five micrograms of protein derived from the cellular extract of a pellet
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43 19 of cultured fibroblasts was subjected to 12.5% sodium dodecyl sulfate polyacrylamide
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45 20 gel electrophoresis (SDS/PAGE). Immunoblotting was performed according to a routine
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47 21 protocol using rabbit polyclonal antibodies against ETF, which were a gift from Dr. T.
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49 22 Hashimoto (Professor Emeritus of Shinshu University, Matsumoto, Japan), and ETFDH,
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51 23 which was purchased from Japan Bio Services Co., Ltd. (Saitama, Japan), as the
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53 24 primary antibodies. Blots were visualized using the Immuno-Pure NBT/BCIP Substrate
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2 1 Kit TM (Promega, Madison WI, USA) [19, 21].
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4 2.7 Gene analysis of *ETFDH* 5

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7 3 Genomic DNA was isolated from fibroblasts using a QIAamp DNA Microkit
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9 4 (QIAGEN GmbH, Hilden, Germany). Each exon of *ETFA*, *ETFB*, and *ETFDH*,
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11 5 including intron/exon boundaries, was PCR-amplified for 30 cycles. Primers for
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13 6 *ETFDH* were prepared as previously reported [2, 14]. The PCR products were purified
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15 7 with a QIAquick PCR Purification Kit (QIAGEN GmbH, Hilden, Germany) and
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17 8 sequenced using an ABI PRISM 310 Genetic Analyzer (PE Applied Biosystems, Foster
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19 9 City, CA, USA) or CEQ 8000 Genetic Analysis System (Beckman Coulter Inc.,
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21 10 Fullerton, CA, USA).
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25 2.8 *In vitro* probe acylcarnitine (IVP) assay 26

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29 12 An IVP assay to evaluate the β -oxidation capacity was performed as previously
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31 13 described [20]. Briefly, confluent cells were harvested by trypsinization and seeded onto
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33 14 6-well microplates with fresh medium (described above) until they again reached
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35 15 confluence. Thereafter, cells were washed twice with D-PBS and cultured at 37°C in 1
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37 16 mL of experimental MEM containing 0.4% essential fatty acid-free BSA, 0.4 mmol/L
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39 17 L-carnitine, and 1% penicillin/streptomycin with 0.2 mmol/L unlabeled palmitic acid.
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41 18 The concentration of ACs in 10 μ L of the culture medium after incubation for 96 hours
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43 19 was determined by MS/MS.
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51 3. Results 52

53 3.1 Urinary organic acid analysis 54

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56 23 No obvious abnormalities were found for urinary OAs under stable conditions
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58 24 for both cases 1 and 2 (Table 2).
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3.2 Blood acylcarnitine analysis

In the AC profiles of blood filter paper, there were no obvious abnormalities in case 1, while there was slight elevation from C4 to C18 in case 2 (Table 3).

In contrast, in the serum AC analysis, slight elevation of C8 and C10 was observed, even under the stable conditions of case 1, and remarkable elevation from C8 to C18 was observed in case 2 (Table 3).

3.3 Histological studies

Muscle stained with Oil-Red O revealed abundant fat deposition in both cases 1 and 2, suggesting metabolic myopathy (Figure 1-A and 1-B).

3.4 Immunoblotting

In both cases 1 and 2, ETFDH protein was not detected, while both ETF α and ETF β proteins were observed to be normal. These findings strongly suggested that both patients had GA2 due to a defect in ETFDH (Figure 2).

3.5 Gene analysis of ETFDH

Mutation analysis revealed that case 1 was a homozygote of 1367C>T (P456L), and case 2 was a compound heterozygote of 890G>T (W297L) and 950C>G (P317R). Eventually, both cases were diagnosed with GA2 due to a defect in ETFDH (Table 2).

3.6 In vitro probe acylcarnitine assay

Only a slight elevation in C10 was observed in case 2, and the elevation of short- to long-chain ACs, which is a characteristic profile for the IVP assay in pediatric cases of GA2, was not observed in either case (Figure 3A, B).

4. Discussion

In this study, we report the clinical, biochemical, and molecular aspects of the

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2 1 adult-onset myopathic form of GA2 in 2 cases. Our cases exhibited the following
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4 2 characteristics compared with pediatric cases: 1) repeated episodes of general fatigue,
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6 3 myalgia, or muscular hypotonia after adulthood (approximately 30 or 40 years of age);
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8 4 2) in routine laboratory findings, slight or moderate elevation of AST, ALT, LDH, and
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10 5 CK; 3) no specific abnormalities for urinary OA analysis under stable conditions; 4) no
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12 6 or barely observable abnormalities in the AC analysis of blood filter paper; 5)
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14 7 significant abnormalities for ACs in the serum; 6) lipid deposition in the muscular
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16 8 biopsy as an initial hint suggesting a GA2 diagnosis; and 7) no abnormalities in the IVP
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18 9 assay for adult-onset cases.
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24 10 In both cases, few or no abnormalities were detected in several examinations,
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26 11 including urinary OA analysis and blood AC analysis. Indeed, cases of adult-onset GA2
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28 12 with little biochemical abnormality have been previously reported [22, 23], suggesting
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30 13 that a biochemical diagnosis of adult-onset GA2 is challenging. Therefore, a number of
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32 14 adult-onset GA2 patients with myopathy of unknown cause might be hidden.
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36 15 Serum AC analysis appeared to be more informative than blood filter paper for
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38 16 diagnosing adult-onset GA2. There are previous reports that serum or plasma AC
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40 17 analysis could be more useful than a dried blood spot for diagnosing long-chain FAODs,
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42 18 such as very long-chain acyl-CoA dehydrogenase deficiency or carnitine
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44 19 palmitoyltransferase-II deficiency [17, 24].
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48 20 The histological findings of lipid deposition provided an initial clue for the
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50 21 diagnosis of GA2 in both of our cases. If fatty degeneration is revealed by muscle
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52 22 biopsy in patients with myopathy of unknown cause, the possibility of FAODs should
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54 23 be considered, even in adult cases.
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58 24 We previously reported that pediatric cases of GA2 could be classified into the
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2 1 severe or milder form using the results of the IVP assay [25]. However, the profiles for
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4 2 the IVP assay in our cases were different from those of the severe or milder forms. In
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6 3 other words, the biochemical characteristics of adult-onset GA2 are different from those
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8 4 of pediatric cases. Additionally, we determined whether abnormal findings in the IVP
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10 5 assay could be improved by bezafibrate [26], but it may be difficult to evaluate the
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12 6 efficacy of bezafibrate for adult-onset GA2 because the profile of the IVP assay in
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14 7 adult-onset GA2 does not encompass specific abnormalities.
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19 8 The clinical findings in case 1 included at least three episodes of
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21 9 unconsciousness, which were estimated to be caused by a hypoglycemic attack.
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23 10 Moreover, the younger brother of case 1 had previously died suddenly from an
24
25 11 unknown cause in his 30s, suggesting that he might also have had GA2 and then
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27 12 developed profound hypoglycemia or arrhythmia, leading to sudden death. There are
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29 13 previous case reports of adult-onset GA2 cases with serious complications, including a
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31 14 25-year-old female who was treated with a ventilator due to respiratory muscle failure
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33 15 [27] and a 19-year-old female patient who had repeated hypoglycemic attacks [28].
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35 16 These cases indicate that critical symptoms can occur in the adult-onset type.
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41 17 Clinical and biochemical features of adult-onset GA2 have recently been
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43 18 reported, as shown in Table 4. All were myopathic cases associated with ETFDH
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45 19 deficiency. However, there is also a report of a late-onset type other than ETFDH
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47 20 deficiency, although this is very rare [29]. The clinical severity varied; severe general
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49 21 symptoms manifested in patients with adult-onset GA2 despite few biochemical
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51 22 abnormalities, as in case 1 reported here and a case reported by Rosenbohm et al. [27],
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53 23 suggesting an unlikely association between the degree of clinical severity and
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55 24 biochemical abnormality.
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1 GA2 has been roughly classified into the neonate-onset and late-onset types [4].

2 However, the clinical course of the “late-onset type” differs substantially among
3 individuals; some cases have encephalopathy or sudden death during the infantile period,
4 while others may only have muscular symptoms in adulthood, as was the case with the
5 patients reported here. Therefore, we propose to distinguish the late-onset type of GA2
6 between the intermediate and myopathic forms, as shown in Table 5, according to the
7 results of the IVP assay as well as age at onset, fatality, and clinical characteristics. The
8 intermediate form (juvenile-onset form) exhibits intermittent attacks, including
9 hypotonia, hypoglycemia, hyperammonemia, and acute encephalopathy-like attack,
10 with typical biochemical abnormalities and relatively high mortality following
11 metabolic stress from an infection or diarrhea in infancy or young childhood. The IVP
12 assay for the intermediate form reveals the elevation of broad ranges in acylcarnitine
13 (C4 to C16) when palmitate is loaded (Figure 3D) [25]. The myopathic form
14 (adult-onset form), in which the patients primarily present with intermittent muscular
15 symptoms after adolescence or adulthood with normal intelligence, offers a favorable
16 life prognosis in many cases. However, it should be noted that muscle symptoms are
17 sometimes exhibited during the infantile period even in the myopathic form [30].

18 The above classification based on the IVP assay can also be used for preclinical
19 risk control of GA2 detected in neonatal mass screening. It is expected that, with the
20 spread of knowledge regarding the clinical characteristics of adult-onset GA2, such a
21 form of GA2 will be found among patients with “myopathy of unknown origin” in the
22 future.

23 24 **Acknowledgements**

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7

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2 **1 Figure legends**
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5 2 Figure 1. Pathological findings from the muscle biopsy (Oil-red O stain).
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7 3 A, case 1, and B, case 2. Arrows indicate lipid deposits.
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11 5 Figure 2. Immunoblots of ETFDH and ETF proteins using fibroblasts.
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13 6 Lanes C1 and C2, normal controls; lanes 1 and 2, cases 1 and 2, respectively. Open and
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17 7 closed triangles represent positive and negative, respectively.
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20 8
21 9 Figure 3. Profiles of the *in vitro* probe assay. Arrows indicate loaded fatty acid (palmitic
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24 10 acid).
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26 11 The Y-axis represents values of acylcarnitines expressed as nmol/mg protein/96 h. A,
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29 12 case 1; B, case 2; C, patient with a severe form of GA2; D, patient with an intermediate
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31 13 form of GA2; and E, healthy controls. Black and white columns indicate our cases and
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34 14 pediatric cases of the severe form, the intermediate form, and the control, respectively.
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1 **Tables**

2 Table 1. Outlines of the patients and results of routine laboratory tests

	Case 1	Case 2	(Reference value*)
Onset Age	40s	31	
Sex	M	M	
Clinical features	myalgia muscle weakness rhabdomyolysis	myalgia muscle weakness	
Routine blood examination			
CBC			
WBC (/ μ L)	4,800	5,000	(3300-8600)
RBC ($\times 10^4$ / μ L)	370	539	(385-438)
Hb (g/dL)	12.3	16.5	(11.0-14.8)
Plt ($\times 10^4$ / μ L)	18.7	20.7	(15.8-35.3)
Biochemical data			
T-Bil (mg/dL)	0.3	0.8	(0.2-1.2)
TP (g/dL)	5.6	7.3	(6.5-8.2)
Alb (g/dL)	3.4	5.1	(3.8-5.1)
AST (IU/L)	<u>197</u>	<u>71</u>	(10-38)
ALT (IU/L)	<u>215</u>	<u>84</u>	(5-40)
LDH (IU/L)	<u>2,903</u>	<u>684</u>	(100-215)
ALP (IU/L)	178	152	(110-340)
CK (IU/L)	<u>2364</u>	<u>689</u>	(36-216)
BUN (mg/dL)	7	10.9	(8.0-21.0)
Cre (mg/dL)	0.35	0.5	(0.44-0.83)
Na (mEq/L)	138	139	(137-146)
K (mEq/L)	3.4	4.1	(3.5-4.9)
Cl (mEq/L)	101	103	(98-109)
Ca (mg/dL)	8.8	10.6	(8.6-10.3)
BS (mg/dL)	90	104	(60-109)

3 * The reference values used at Shimane University.

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1 Table 2. Results of special examinations

	Case 1	Case 2
Muscle biopsy	<i>lipid deposit</i>	<i>lipid deposit</i>
Urinary organic acid analysis	Normal	non-specific finding
Blood acylcarnitine analysis (dried blood spot)	Normal	
Gene analysis of <i>ETFDH</i>	1367C>T (P456L) (homozygote)	890G>T (W297L)/ 950C>G (P317R)

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4 Table 3. Comparison of free carnitine and acylcarnitine in DBS and serum

	Dried blood spot			Serum		
	Case 1	Case 2	(Reference)	Case 1	Case 2	(Reference)
C0	37.94	45.37	(20 - 60)	32.79	52.35	(10 - 55)
C2	28.07	46.19	(5 - 45)	11.56	33.02	(4 - 60)
C4	0.37	<u>1.77</u>	(<1.4)	0.27	0.78	(<1.65)
C8	0.06	<u>0.98</u>	(<0.25)	<u>1.92</u>	<u>1.61</u>	(<0.46)
C10	0.18	<u>2.03</u>	(<0.35)	<u>1.88</u>	<u>4.63</u>	(<0.8)
C12	0.09	<u>0.8</u>	(<0.4)	0.24	<u>1.35</u>	(<0.4)
C14	0.38	<u>1.01</u>	(<0.7)	0.08	<u>3.29</u>	(<0.3)
C16	2.90	3.12	(<7.0)	0.22	<u>1.19</u>	(<0.5)
C18	1.14	<u>2.32</u>	(<2.1)	0.06	<u>0.55</u>	(<0.3)

5 The reference values reported here are those used at Shimane University. Values judged

6 as abnormal are underlined.

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1 Table 4. Recently reported clinical and biochemical features for adult-onset GA2

No	Sex	Age at onset (year)	Myalgia	Muscle weakness	Other symptoms	Laboratory data			Increased urinary organic acid	Elevated acylcarnitines		Gene	Gene mutation		Reference
						Elevated trans-aminase	LDH (IU/L)	CK (IU/L)		DBS	Serum (Plasma)		Allele 1	Allele 2	
Our cases															
1	M	40s	+	+	coma	+	2,903	3,000	normal	normal	C8-C10	<i>ETFDH</i>	P456L	P456L	our case
2	M	31	+	+	no	+	2,860	1,897	normal	C4-C12	C8-C18	<i>ETFDH</i>	W297L	P317R	our case
Previously reported cases															
3	M	42	+	+	no	N/A	942	1,855	GA, 2HG, EMA	C4, C5, C8, C10, C14	N/A	<i>ETFDH</i>	I243T	T294I	Köppel et al, 2006 [10]
4	F	24	+	+	no	N/A	N/A	677	N/A	C8-C12	N/A	<i>ETFDH</i>	L409F	V291G	Wen et al, 2010 [23]
5	F	23	+	+	vomiting	N/A	N/A	513	N/A	C8-C12	N/A	<i>ETFDH</i>	L409F	V291G	Wen et al, 2010 [23]
6	F	48	+	+	vomiting	N/A	N/A	128	N/A	C0 (↓), C8-C10	N/A	<i>ETFDH</i>	Y257C	not detected	Wen et al, 2010 [23]
7	F	22	-	+	no	N/A	N/A	478	GA, 2HG, EMA, DCA, KB	C4-OH, C10-C14	N/A	<i>ETFDH</i>	Y257C	V291G	Wen et al, 2010 [23]
8	F	33	+	+	no	N/A	N/A	352	GA, 2HG, EMA, DCA, KB	C0 (↓), C12-C14	N/A	<i>ETFDH</i>	Y257C	325del48	Wen et al, 2010 [23]
9	F	63	-	+	no	N/A	N/A	2,120	GA, 2HG, EMA, DCA	C0, C5-C14	N/A	<i>ETFDH</i>	N/A heterozygote?	none	Wen et al, 2010 [23]
10	F	23	+	+	vomiting	N/A	N/A	1,998	GA, 2HG, EMA, DCA, KB	C8-C14	N/A	<i>ETFDH</i>	M404T	not detected	Wen et al, 2010 [23]
11	F	22	+	-	no	N/A	N/A	339	normal	C0	N/A	<i>ETFDH</i>	L409F	not detected	Wen et al, 2010 [23]
12	M	46	+	+	difficulty in breathing	+	543	5,995	GA, 2HG, DCA	N/A	N/A	<i>ETFDH</i>	M404T	D596N	Izumi et al, 2011 [11]
13	F	55	+	+	no	N/A	N/A	8,000	normal	N/A	C4-C18	<i>ETFDH</i>	H293D	not detected	Kaminsky et al, 2011 [22]
14	M	36	-	-	exercise intolerance	N/A	1,161	3,055	2HG, 2-OH adipate	N/A	N/A	<i>ETFDH</i>	D511N	W603X	Sugai et al, 2012 [12]
15	M	53	+	+	osphyalgia, nausea	+	600	571	GA, 2HG, EMA	C8-C12	N/A	<i>ETFDH</i>	P508T	N528KfsX3	Zhao et al, 2012 [13]
16	F	24	+	+	vomiting, respiratory insufficiency	+	N/A	20,000	2HG, EMA, DCA, HG, SG	N/A	C2 (↓), C14:1	<i>ETFDH</i>	S515I	S515I	Rosenbohm et al, 2014 [27]

1 LDH: lactate dehydrogenase, CK: creatine kinase, DBS: dried blood spot, N/A: not available, GA: glutarate, HG: 2-hydroxyglutarate,
 2 EMA: ethylmalonate, DCA: dicarboxylate, KB: ketone body, HG: hexanoylglycine, SG: suberylglycine, and (↓): decreased

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Table 5. Classification of glutaric acidemia type II based on the severity and IVP assay results

Clinical form	Age at onset	Clinical course	Mortality	Biochemical abnormality	In vitro probe assay with C16 loaded
1. Severe form (neonatal-onset)	soon after birth	rapid onset and early death after birth hyperammonemia, hypoglycemia, or cardiomyopathy	++	++	marked elevation of C16
2. Intermediate form (juvenile-onset)	infantile or childhood	episodes of lethargy, liver dysfunction, or hypoglycemia occasionally encephalopathy or even sudden death	+	+	elevation of C4 to C16
3. Myopathic form (adult-onset)	school-age or adulthood	episodes of myalgia, muscle weakness, fatigue, or liver dysfunction	-	±	almost normal

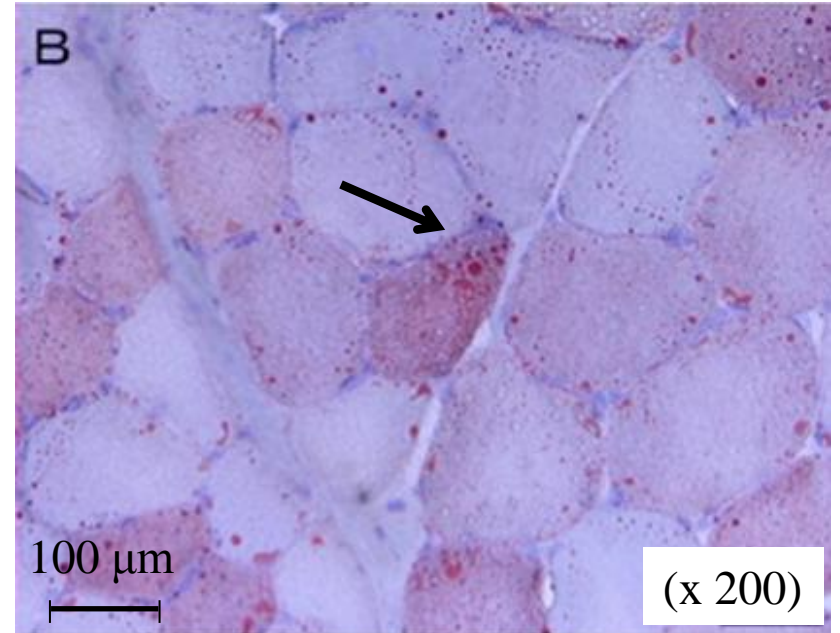
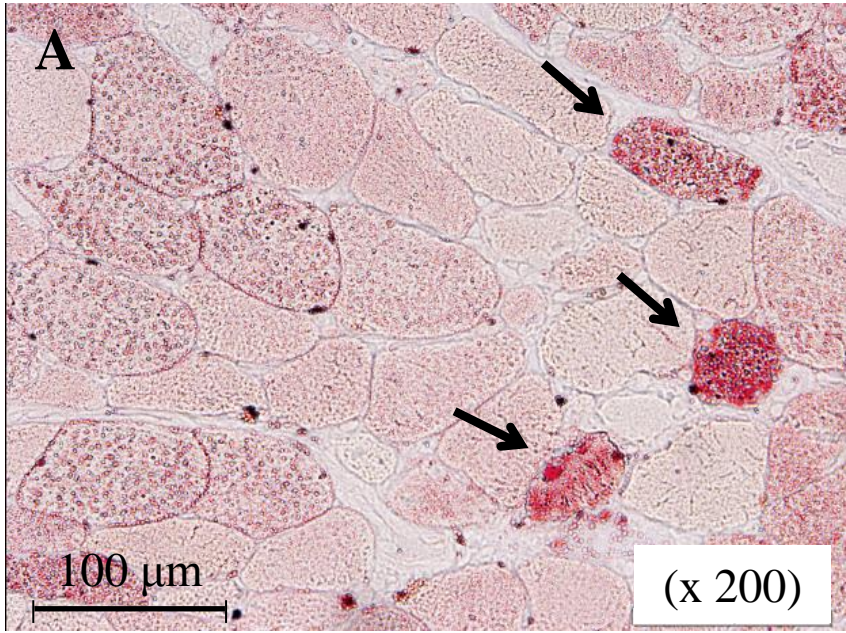


Figure 1. Pathological findings of muscle biopsy (oil red stain)

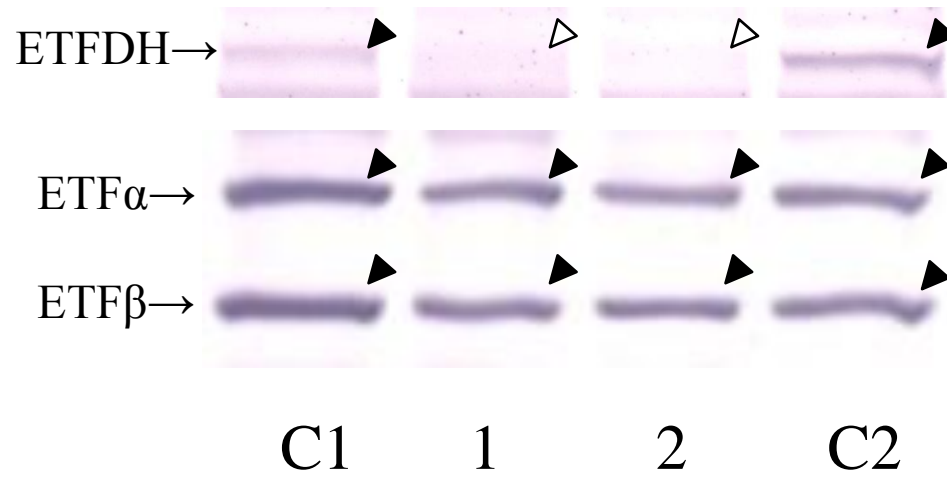
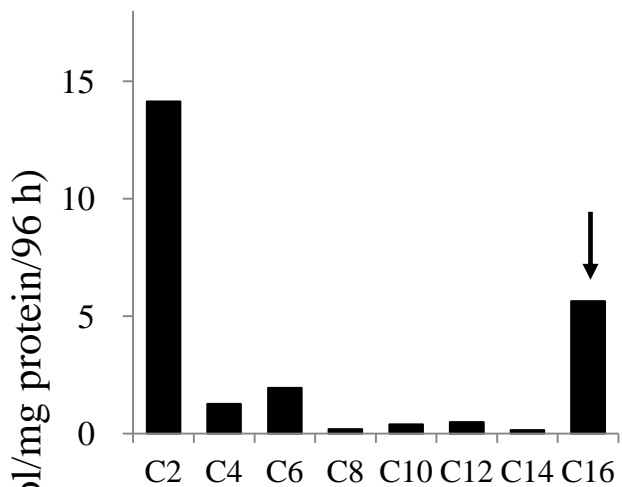
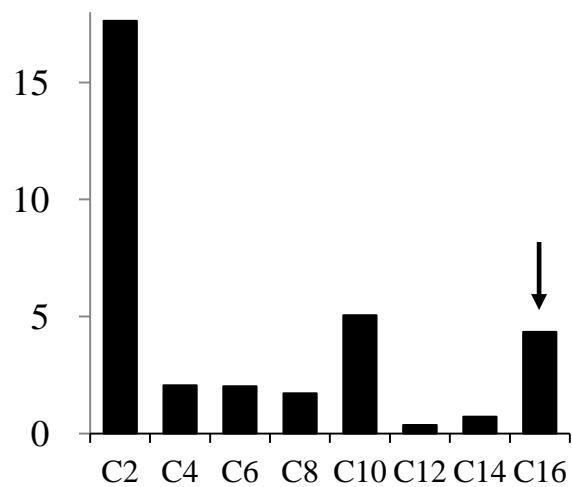


Figure 2. Immunoblotting of fibroblasts

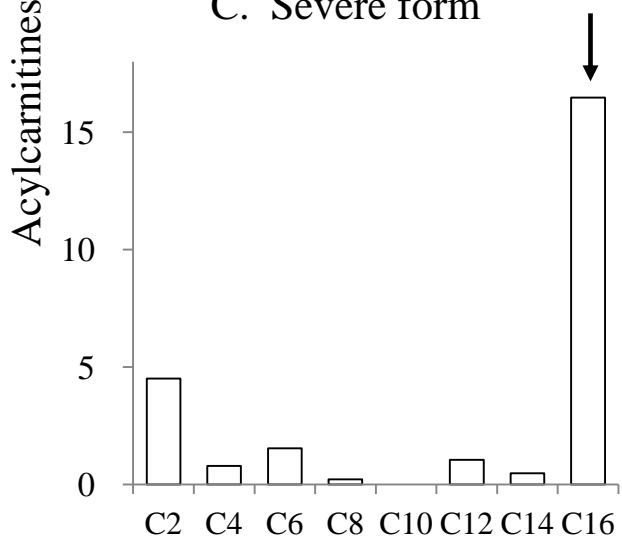
A. Case 1



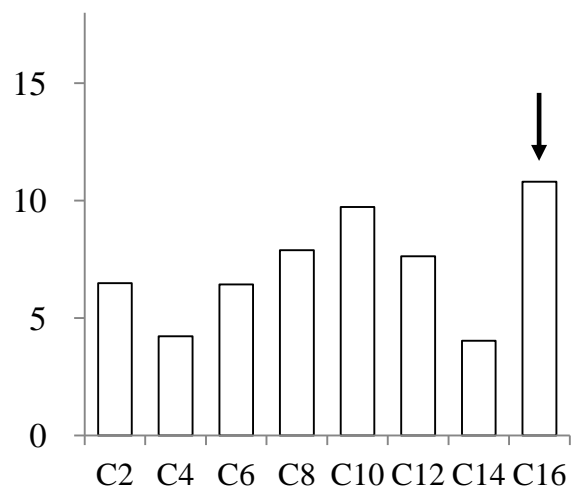
B. Case 2



C. Severe form



D. Intermediate form



E. Control

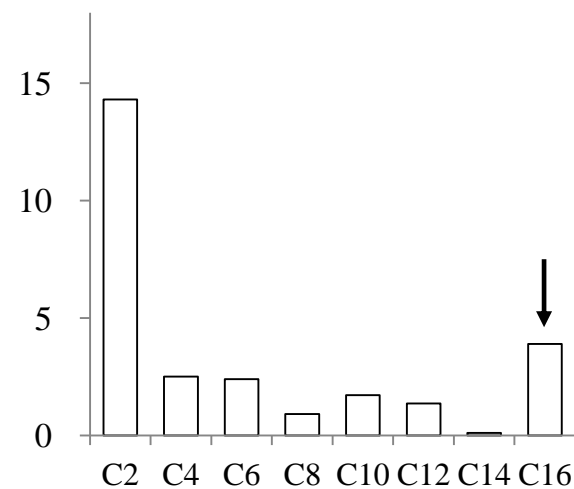


Figure 3. Profiles of *in vitro* probe assay in GA2